

T. Kolev,<sup>a</sup> M. Spitteller,<sup>b</sup> T. van Almsick,<sup>c</sup> H. Mayer-Figge<sup>c</sup> and W. S. Sheldrick<sup>c\*</sup><sup>a</sup>Institute of Organic Chemistry, Bulgarian Academy of Sciences, Acad. G. Bonchev Street, Building 9, 1113 Sofia, Bulgaria, <sup>b</sup>Institut für Umweltforschung, Universität Dortmund, Otto-Hahn-Strasse 6, D-44221 Dortmund, Germany, and <sup>c</sup>Lehrstuhl für Analytische Chemie, Ruhr-Universität Bochum, Universitätsstrasse 150, 44780 Bochum, GermanyCorrespondence e-mail:  
william.sheldrick@rub.de

## Key indicators

Single-crystal X-ray study  
*T* = 294 K  
Mean  $\sigma(\text{C}-\text{C}) = 0.008 \text{ \AA}$   
*R* factor = 0.067  
*wR* factor = 0.172  
Data-to-parameter ratio = 13.6For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

## Benzamidinium dihydrogenphosphate

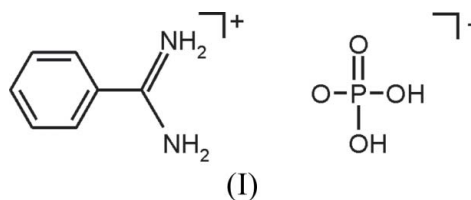
Cations and anions of benzamidinium dihydrogenphosphate,  $\text{C}_7\text{H}_9\text{N}_2^+ \cdot \text{H}_2\text{PO}_4^-$ , are connected by  $\text{NH}_2 \cdots \text{O}=\text{P}$  and  $\text{NH}_2 \cdots \text{O}(\text{H})-\text{P}$  hydrogen bonds between benzamidinium  $\text{NH}_2$  groups and dihydrogenphosphate O atoms into an infinite three-dimensional network. The anions are connected through  $\text{P}-\text{OH} \cdots \text{O}=\text{P}$  interactions.

Received 24 January 2007

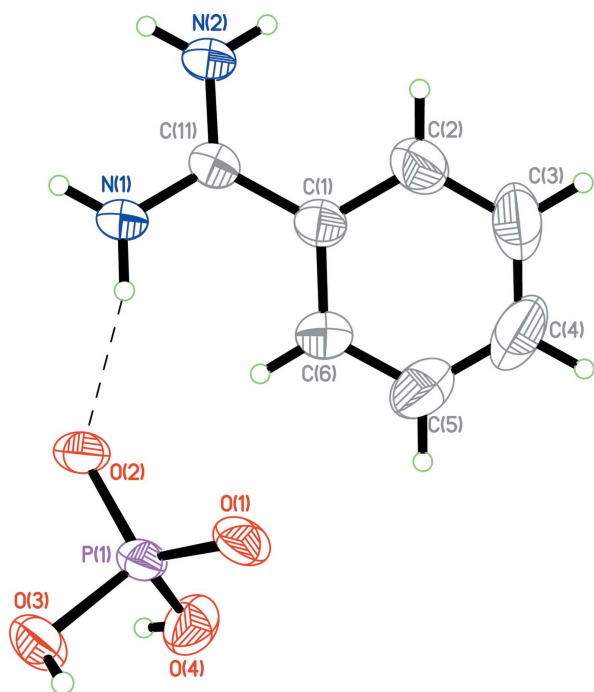
Accepted 24 January 2007

## Comment

The structure of benzamidinium dihydrogenphosphate, (I), has been determined as a part of our continuing synthetic, spectroscopic and structural investigations of dihydrogenphosphates (Kolev *et al.*, 2007) and hydrogensquarates (Kolev *et al.*, 2005; Kolev, Spitteller, *et al.*, 2006; Kolev, Yancheva *et al.*, 2006) of biologically active amines and various substituted amidines. It is known that factor Xa (FXa) is a key enzyme for the intervention of the blood coagulation cascade and for the development of new antithrombotic agents (Willardsen *et al.*, 2004). Many drugs developed as potential inhibitors of coagulation enzymes contain the benzamidine functionality (Koshio *et al.*, 2005). Commonly, compounds of this class are reversible inhibitors of trypsin and other peptidases that show a selectivity for arginine, or possibly lysine. The cationic amidino group of the inhibitor interacts with a carboxylate located at the bottom of the S1 subsite, and there are also hydrophobic interactions with the sides of the S1 pocket. Benzamidine is, therefore, a common component of mixtures of inhibitors designed to suppress peptidase activity in biological samples (Bode *et al.*, 1983; Krieger *et al.*, 1974; Powers & Harper, 1986). Its cation has also been included in a number of protein structure determinations (Bode *et al.*, 1990; Marquart *et al.*, 1983; Sprang *et al.*, 1987).

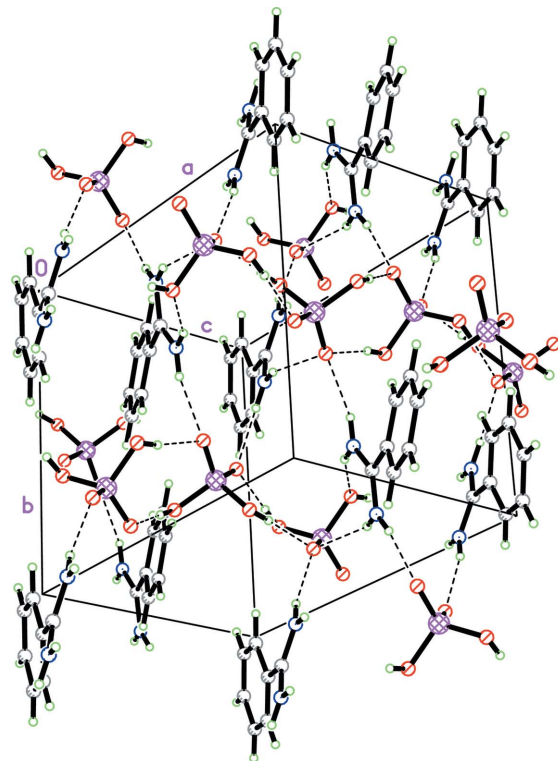


Phosphates are an essential nutrient for all organisms, which have, therefore, evolved regulatory mechanisms for its acquisition, storage and release (Torriani-Gorini *et al.*, 1994). The evidence, to date, indicates that regulation of the overall renal tubular phosphate transfer occurs at the level of the proximal tubular border membrane Na/phosphate cotransport system (Dousa, 1996; Kempson, 1996). Hyperphosphatemia and increased calcium phosphate production are important



**Figure 1**

The asymmetric unit of (I), showing the atom-labelling scheme. Displacement ellipsoids are drawn at the 50% probability level. The dashed line indicates a hydrogen bond.



**Figure 2**

Linkage of the anions and cations of (I) into a three-dimensional network through hydrogen bonds (dashed lines).

contributors to vascular calcifications in patients with uremia (Block *et al.*, 1998).

A detailed knowledge of the structure and spectroscopic properties of the benzamidinium cation is a prerequisite for understanding its binding properties and (I) contains this cation in the presence of the biologically important dihydrogenphosphate anion. The crystal structure of benzamidinium has already been reported (Barker *et al.*, 1996).

The asymmetric unit of (I) is depicted in Fig. 1 and a packing diagram in Fig. 2. N—H...O hydrogen bonds between the benzamidinium amino groups and the dihydrogenphosphate counter-ions link the cations and anions of (I) into a three-dimensional network. The anions are connected into infinite double chains through O—H...O interactions running along the *c* axis.

## Experimental

The starting compound benzamidinium was received as a white powder from Bachem (Switzerland) and recrystallized from methanol. A high yield and suitable crystals of (I) for X-ray analysis were obtained by mixing a 20 ml methanol solution of benzamidinium (6 mmol, 776 mg) with an equimolar amount of 50% phosphorous acid. The reaction mixture was stirred for 10 h at room temperature and product formation monitored by thin-layer chromatography. After completion of the reaction, the resulting solution was filtered off and the filtrate set aside, affording crystals of (I) after 24 h. The product was separated by filtration, dried in air and characterized by elemental analysis, mass spectrometry and IR, Raman and NMR spectroscopy. The IR spectrum of (I) exhibits  $\nu$  N<sup>+</sup>H<sub>2</sub> stretching modes between 3400 and 3000 cm<sup>-1</sup>, with the highest frequency peak at 3353 cm<sup>-1</sup>.

This region overlaps with the  $\nu$  OH bands of hydrogen-bonded dihydrogenphosphate anions. The intense peaks at 1231 and 1088 cm<sup>-1</sup> can be assigned to  $\nu$  P=ON and  $\nu$  PO(H), respectively, and the peaks at 1700 and 1680 cm<sup>-1</sup> to bending  $\delta$  N<sup>+</sup>H<sub>2</sub> modes. A full theoretical vibrational analysis and experimental assignment by means of solid-state linear polarized IR spectroscopy is now in progress for (I) and will be published at a later date.

### Crystal data

C<sub>7</sub>H<sub>9</sub>N<sub>2</sub><sup>+</sup>·H<sub>2</sub>O<sub>4</sub>P<sup>-</sup>  
*M<sub>r</sub>* = 218.15  
 Monoclinic, *P*2<sub>1</sub>/*c*  
*a* = 12.259 (3) Å  
*b* = 10.067 (2) Å  
*c* = 8.0578 (16) Å  
 $\beta$  = 91.28 (3)°  
*V* = 994.2 (3) Å<sup>3</sup>

*Z* = 4  
*D<sub>x</sub>* = 1.457 Mg m<sup>-3</sup>  
 Mo K $\alpha$  radiation  
 $\mu$  = 0.27 mm<sup>-1</sup>  
*T* = 294 (2) K  
 Broken prism, colourless  
 0.49 × 0.30 × 0.23 mm

### Data collection

Siemens P4 four-circle diffractometer  
 $\omega$  scans  
 Absorption correction:  $\psi$  scan (XPREP; Sheldrick, 1995)  
*T<sub>min</sub>* = 0.877, *T<sub>max</sub>* = 0.939  
 1875 measured reflections

1739 independent reflections  
 1071 reflections with *I* > 2 $\sigma$ (*I*)  
*R<sub>int</sub>* = 0.055  
 $\theta_{\max}$  = 25.0°  
 3 standard reflections every 100 reflections  
 intensity decay: 2%

### Refinement

Refinement on *F*<sup>2</sup>  
*R* [*F*<sup>2</sup> > 2 $\sigma$ (*F*<sup>2</sup>)] = 0.067  
*wR* (*F*<sup>2</sup>) = 0.172  
*S* = 1.07  
 1739 reflections  
 128 parameters  
 H-atom parameters constrained

$w = 1/[\sigma^2(F_o^2) + (0.0866P)^2]$   
 where  $P = (F_o^2 + 2F_c^2)/3$   
 $(\Delta/\sigma)_{\max} < 0.001$   
 $\Delta\rho_{\max} = 0.40$  e Å<sup>-3</sup>  
 $\Delta\rho_{\min} = -0.38$  e Å<sup>-3</sup>  
 Extinction correction: SHELXL97  
 Extinction coefficient: 0.007 (2)

**Table 1**

Hydrogen-bond geometry (Å, °).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
N1—H12 $\cdots$ O2 <sup>i</sup>	0.86	2.06	2.891 (5)	161
N1—H11 $\cdots$ O3 <sup>ii</sup>	0.86	2.22	3.028 (5)	157
N2—H22 $\cdots$ O1 <sup>iii</sup>	0.86	2.01	2.833 (5)	160
N2—H21 $\cdots$ O2 <sup>ii</sup>	0.86	2.12	2.892 (5)	149
O3—H3' $\cdots$ O2 <sup>iv</sup>	0.82	1.83	2.643 (4)	171
O4—H4' $\cdots$ O1 <sup>v</sup>	0.82	1.80	2.616 (4)	178

Symmetry codes: (i)  $-x + 2, y + \frac{1}{2}, -z + \frac{1}{2}$ ; (ii)  $x + 1, y, z$ ; (iii)  $-x + 2, -y, -z$ ; (iv)  $x, -y + \frac{1}{2}, z - \frac{1}{2}$ ; (v)  $x, -y + \frac{1}{2}, z + \frac{1}{2}$ .

H atoms were treated as riding, with C—H = 0.93, N—H = 0.86 and O—H = 0.82 Å and with  $U_{iso}(H) = 1.2U_{eq}(C,N)$  and  $1.4U_{eq}(O)$ .

Data collection: *R3m/V* (Siemens, 1989); cell refinement: *R3m/V*; data reduction: *XDISK* (Siemens, 1989); program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *SHELXTL-Plus* (Sheldrick, 1995); software used to prepare material for publication: *SHELXL97*.

TK and MS thank the DAAD for a grant within the priority programme 'Stability Pact South-Eastern Europe', and the Alexander von Humboldt Foundation.

**References**

Barker, J., Phillips, P. R., Wallbridge, M. G. H. & Powell, H. R. (1996). *Acta Cryst.* **C52**, 2617–2619.  
 Block, G. A., Hulbert-Shearon, T. E., Levin, N. W. & Port, F. K. (1998). *Am. J. Kidney Dis.* **31**, 607–617.

Bode, W., Chen, Z., Bartels, K., Kutzbach, C., Schmidt-Kasner, G. & Bartunik, H. (1983). *J. Chem. Biol.* **164**, 237–241.  
 Bode, W., Turk, D. & Stürzebecher, J. (1990). *Eur. J. Biochem.* **193**, 175–180.  
 Dousa, T. (1996). *Kidney Int.* **49**, 997–1004.  
 Kempson, S. (1996). *Kidney Int.* **49**, 1005–1009.  
 Kolev, T., Spitteller, M., Sheldrick, W. S. & Mayer-Figge, H. (2005). *Acta Cryst.* **E61**, o4292–o4294.  
 Kolev, T., Spitteller, M., Sheldrick, W. S. & Mayer-Figge, H. (2006). *Acta Cryst.* **C62**, o299–o300.  
 Kolev, T., Spitteller, M., van Almsick, T., Sheldrick, W. S. & Mayer-Figge, H. (2007). *Acta Cryst.* **E63**, o179–o181.  
 Kolev, T., Yancheva, D., Spitteller, M., Sheldrick, W. S. & Mayer-Figge, H. (2006). *Acta Cryst.* **E62**, o463–o465.  
 Koshio, H., Hirayama, F., Ishihara, T., Shiraki, R., Shigenaga, T., Taniuchi, Y., Sato, K., Moritani, Y., Iwatsuki, Y., Kaku, S., Katayama, N., Kawasaki, T., Matsumoto, Y., Sakamoto, S. & Tsukamoto, S. (2005). *Bioorg. Med. Chem.* **13**, 1305–1323.  
 Krieger, M., Kay, L. M. & Stroud, R. M. (1974). *J. Mol. Biol.* **83**, 209–230.  
 Marquart, M., Walter, J., Deisenhofer, J., Bode, W. & Huber, R. (1983). *Acta Cryst.* **B39**, 480–490.  
 Powers, J. C. & Harper, J. W. (1986). *Proteinase Inhibitors*, edited by A. J. Barrett & G. Salvesen, pp. 55–152. Amsterdam: Elsevier.  
 Sheldrick, G. M. (1995). *SHELXTL-Plus*. Release 5.03 for the Siemens R3 crystallographic research system. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.  
 Sheldrick, G. M. (1997). *SHELXS97* and *SHELXL97*. University of Göttingen, Germany.  
 Siemens (1989). *R3m/V Crystallographic-Systems User's Guide*. Version 3.2. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.  
 Sprang, S., Standing, T., Fletterick, R. J., Strong, R. M., Finer-Moor, J., Xuong, N. H., Hamlin, N. R., Rutter, W. & Kreik, C. S. (1987). *Science*, **237**, 905–909.  
 Torriani-Gorini, A., Silver, S. & Yagil, E. (1994). *Phosphate in Microorganisms: Cellular and Molecular Biology*. Washington, DC: American Society for Microbiology.  
 Willardsen, J. A., Dudley, D. A., Cody, W. L., Chi, L., McClanahan, T. B., Mertz, T. E., Potoczak, R. E., Narasimhan, L. S., Holland, D. R., Rapundalo, S. T. & Edmunds, J. J. (2004). *J. Med. Chem.* **29–47**, 4089–4099.